Volume 6, Number 1, December 2016

Website: http://jsci.utq.edu.iq

Email: utjsci@utq.edu.iq

Molecular characterization of mecA gene in Methicillin-Resistant Staphylococcus aureus

Saad Salman Hamim

Department of Pathological Analysis- College of Science- University of Thi-Qar- Iraq

E-mail: hamim_pa@sci.utq.edu.iq Tel: 009647801016056

Abstract:

Background & objectives: It is clear now that methicillin-resistant *Staphylococcus aureus* (MRSA) strains considered as one of the most bacteria responsible for different diseases among humans and animals. The present study aimed to detect the molecular profile of methicillin-resistant *S. aureus* isolated from skin abscess patients in Nassyriah City, Southern Iraq.

Methods: During the period of from June 2014 to February 2015, 120 *S. aureus* were isolated from abscess patient in two governmental hospitals, and subjected to conventional Polymerase Chain Reaction which used for the amplification of 310 bp *mecA* gene. Three PCR products of *mecA* were named primarily (THQR1, THQR2, and THQR3) were selected and subjected to partial DNA sequencing for the *mecA* gene to follow up their possible relationship between these local isolates and what recorded globally in Genbank. **Results:** Only 64 *S. aureus* isolates were diagnosed phenotypically as MRSA (53.33%), and 88/120 (73.33%) of *S. aureus* were positive for the targeted gene. The Three PCR products of *mecA* were registered in Genbank under the official accession numbers of (KY468502, KY468503 and KY468504, respectively). The constructed phylogenetic tree showed that *S. aureus* KY468502 and KY468504 were highly relative to each other in comparison with *S. aureus* KY468503 that revealed a close relatedness to *S. aureus* TN/CN/1/12, *mecA* gene beta-lactam, and *mecA* gene isolated in USA.

Interpretation & conclusions: The present study results confirmed the importance of *mecA* gene in MRSA detection and highlighted the increasing manner of its prevalence in Iraq, furthermore, the importance of molecular techniques as an epidemiological tool.

Key words: Gene sequencing; Methicillin-resistant; Staphylococcus aureus; Phylogenetic tree; mecA gene of S. aureus

الخصائص الجزيئية لجين mecA في بكتريا المكورات العنقودية الذهبية المقاومة للمتسيلين

الخلاصة:

الخلفية والأهداف: أصبح واضحا الآن أن المكورات العنقودية المقاومة للمشيلين تعتبر من أكثر أنواع البكتريا المسئولة عن مختلف الأمراض للإنسان والحيوانات. هدفت الدراسة الحالية للكشف عن الخصائص الجزيئية لبكتريا المكورات العنقودية المقاومة للمشيلين المعزولة من مرضى الخراج الجلدي في مدينة الناصرية , جنوب العراق.

طرق العمل: خلال الفترة من شهر حزيران 2014 إلي شباط 2015 تم عزل 120 عزلة من بكتريا المكورات العنقودية من مرضي الخراج في انتين من المستشفيات الحكومية وقد تم إخضاعها لتقنية تفاعل السلسلة المتبلمرة التقايدي والذي استخدم لتضخيم الجين *meo*A وبطول 310 زوج قاعدي. تمت

Website: http://jsci.utq.edu.iq

Email: utjsci@utq.edu.iq

Volume 6, Number 1, December 2016

التسمية الأولية لثلاثة من نواتج التضخيم للجين بـ THQR1 و THQR3 وتم استخدامها لحساب التسلسل الجيني الجزئي للجين لأجل متابعة علاقاتها المحتملة مع ما مسجل عالميا في بنك الجينات.

النتائج: تم تشخيص 64 عزلة من بكتريا المكورات العنقودية الذهبية فقط مظهريا على أنها مقاومة للمثيسيلين (53,33%) وكانت 88 /120 قد أظهرت كشفا موجبا للجين mecA (73,33 %). تم تسجيل نواتج الترحيل الثلاث للجين في بنك الجينات تحت أرقام الانضمام الرسمية KY468502 و KY468503 و KY468504 على التوالي. أظهرت الشجرة التطورية التي تم بناؤها أن بكتريا *S. aureus للس*لالتين KY468502 و TN/CN/1/12 و كانتا ذات علاقة تطورية متقاربة لبعضهما مقارنة مع السلالة KY468503 التي أظهرت تقاربا مع بكتريا *S. aureus ليران الحري*

التفسيرات والاستنتاجات: أكدت نتائج الدراسة الحالية على أهمية استخدام الجين *mec*A في الكشف عن المكورات العنقودية الذهبية المقامة للمثسيلين وتم تسليط الضوء على الزيادة المطردة لانتشارها في العراق , بالإضافة إلى التأكيد على أهمية التقنيات الجزيئية كأدوات للدراسات الوبائية.

Introduction:

Staphylococcus aureus is usually a member of normal skin flora and the nasal cavity, almost can be responsible for different superficial and deep skin infections; bones; heart; and lung infections (Schito, 2006). In the early of 1960_s ; semi-synthetic β -lactam resistant pencillins, such as mecithillin and oxacillin were introduced, and declining of multi-drug resistance S. aureus was noticed clearly (Shanson, 1981). Unfortunately, after about one decade later, strains resistant to these pencillins, especially, methicillinresistance of S. aureus (MRSA) emerged with increased manner (Jensen and Lyon, 2009). Nowadays, MRSA became as one of the most potential pathogen worldwide with the emergence of community acquired (CA-MRSA) and hospital acquired (HA-MRSA) strains. As a fact accompli, it is difficult to differentiate between these two MRSA types, since CA-MRSA could spread into hospitals (Jarvis et al., 2007; Wannet et al., 2003). The molecular origin of methicillin resistant is due to the presence of mecA gene, which is a part of Staphylococcal cassette chromosome mec (SCC_{mec}) . About eleven different SCC_{mec} types are reported, and continue to be used in the classification of MRSA strains (Rahimi et al., 2014 ; Rahimi and Karimi, 2015). The current study designed to determine the molecular profile of mecA gene among MRSA isolated from skin abscess patients in Nassyriah city, Iraq.

Materials and Methods:

Samples collection: A total of 120 S. aureus isolates were used in the present study which randomly

collected from outpatients with skin abscesses in two governmental hospitals during the period from June 2014 to February 2015, in Nassyriah city, Iraq.

Laboratory methods: All *S. aureus* isolates used in this study were taken from pus swabs and diagnosed depending on Gram's stain; cultural characteristic on Blood agar (BA) base and Manitol salt agar (MSA), followed by conventional biochemical tests (Harley and Prescott, 2002). The diagnosis was confirmed by API system (BioMerieux/France).

Antibiotic susceptibility test for MRSA isolates: All MRSA isolates were subjected to antibiotic susceptibility by using disc diffusion method as mentioned by(Bauer *et al.*, 1966), with $5\mu g$ of methicillin and 10 μg oxacillin (Bioanalyse, Turkey). The inhibition zone diameters were measured and interpreted according to (CLSI, 2009).

Detection of mecA gene by Polymerase Chain Reaction: S. aureus previously extracted DNA was used for the amplification of mecA gene. A volume of 20 µl PCR reaction mixture consisting of 10 µl master mix, 1.25 µl of each forward and reverse primers specific for the target gene, 5 µl of DNA template, and the volume was completed by adding nuclease free water. A 310-bp fragment of the mecA was amplified primers; mecA-F: 5' GTA GAA ATG ACT GAA CGT CCG ATA A -3' and mecA-R: 5' CCA ATT CCA CAT TGT TTC GGT CTA A -3'. The mixture was briefly centrifuged and the tubes was transferred into PCR apparatus (ESCO, India) which has been programmed with the following conditions: an initial denaturation step for 4 minutes at 94°C with one cycle, 30 cycles of amplification were performed as follows: denaturation

Website: http://jsci.utq.edu.iq

Email: utjsci@utq.edu.iq

Volume 6, Number 1, December 2016

at 94°C for 45 seconds, annealing at 50°C for 45 seconds and extension at 72°C for 1 minute, followed by a final extension step at 72°C for 2 minutes (Jonas *et al.*, 2002).

DNA sequencing: Three PCR products of *mecA* genes that represent methicillin-resistant *S. aureus* strains, were selected for sequencing, and forward and reverse primers for the target gene were sent outside Iraq to be sequenced (Macrogen, Korea). Basic Local Alignment Search Tool analysis (BLAST) was lead to blast algorithm. The samples sequences which designated as (THQR1, THQR2, and THQR3) were edited, aligned, and compared with the reference sequences using BioEdit sequence Alignment Editor Software Version 7.1 (DNASTAR, USA) (Hall, 1999). A phylogenetic tree for each gene sequence was constructed by using MEGA7 software (Kumar *et al.*, 2016).

Results:

Phenotypic characterization of methicillin-resistant *Staphylococcus aureus* (MRSA): Out of 120 MSA positive *S. aureus* isolated from skin abscess patients, 64 isolates showed resistant to methicillin and 120 to oxacillin discs which represent a resistant rate of 53.33% and 100%, respectively.

Detection of *mecA* **gene**: All *S. aureus* isolates were subjected to molecular detection of *mecA* gene. Among the assayed *S. aureus*, 88/120 amplified the targeted gene, which represent a percentage of 73.33%, with a molecular weight of approximately 310 bp (Figure 1).

DNA sequencing: The three selected phenotypic and molecular MRSA isolates were subjected to partial DNA sequencing for mecA gene. A FASTA format files containing the local strains sequences were used to assess a molecular relationship between Nassyriah city, Southern Iraq, isolates and other global sequences to find out the possible closely related strains. S. aureus THQR1, THQR2, and THQR3 isolates (with a query lengths of 253, 320, and 277 nucleotide, respectively), showed a phylogeny percentage of 99% when compared by BLAST algorithm. The Three PCR products of mecA S. aureus isolates were granted Genbank accession numbers of (KY468502, KY468503 and KY468504, respectively). A phylogenetic tree with all sequences were constructed. Figure 2, illustrates the maximum likelihood tree for the locally three S. aureus MRSA isolates when aligned with similar sequences around the world. The analysis involved four nucleotide sequences of S. aureus mecA genes which were under the following accession numbers: TN/CN/1/12, mecA gene beta-lactam, mecA gene, and NCTC8325, in which all of them were isolated in USA. A different homology and similarities were noticed between the local MRSA isolates and the American ones.



Fig. 1: PCR amplification of 310 bp *mec*A gene by 1.4% agarose gel electrophoresis, where M: ladder, lane(1-9): positive results. Lane (10 and 11): negative.



Fig. 2: A dendrogram showing the neighbor joining phylogenetic tree of Thi-Qar *mecA* isolates and the related strains from Genbank.

Discussion

According to the growing evidences which considered methicillin-resistant *Staphylococcus aureus* (MRSA), as one of the most significant pathogen that has emerged in the last four decades, a global consensus was assessed about the medical importance of MRSA detection for both patients care and a proper use of infection control resources (AL-Ruaily and Khalil, 2011). As a whole comparison between phenotypic and PCR-based methods for detection of MRSA, and as shown in the present study results, it is still leaning towards the sensitivity of molecular techniques.

Website: http://jsci.utq.edu.iq

Email: utjsci@utq.edu.iq

Volume 6, Number 1, December 2016

However, an important opinion indicates that methicillin-resistant can be due to not only to the presence of the mecA gene alone; but by a cluster of this gene and *ica* gene (Memmi et al., 2008). A lot of local similar studied targeted mecA gene, as a molecular indicator of MRSA in different parts of Iraq, with variable S. aureus samples. A study in Kurdstan region, Northern Iraq, recorded a MRSA prevalence of 51.7% (Hussain, 2016). Other study in Iraq, revealed a slightly, high MRSA rates with 93.4% and 73.2% for Baghdad and Wasit Provinces, respectively (Al-Dahbi and Al-Mathkhury, 2013 ; Al-Mayahie et al., 2015). According to the phenotypic resistant, a study in Thi-Oar Province, revealed a MRSA occurrence of 67.46% (Degaim et al., 2015); which seems to be higher than the results obtained by the present study. Methicillin resistant rates continue to be confusing when highlighted the similar studies conducting in neighboring countries, such as what recorded in Saudi Arabia and Iran with a MRSA rates of 86.7% and 56.5%, respectively (AL-Ruaily and Khalil, 2011; Ghasemiam and Mirzaee, 2016). The differences of MRSA prevalence among the regions in Iraq, or even worldwide, may be explained by the variation of geographical distribution, S. aureus sample sources, and types and accuracy of techniques used. The sequence types of the locally targeted S. aureus KY468502 and KY468504 isolates seems to be more relative, since they originate from a single ancestral root and branched as a monophyletic sister cladding form. However, these two isolates were relatively, far from S. aureus KY468503, which was closely related to S. aureus TN/CN/1/12, mecA gene beta-lactam, and mecA gene isolates, followed by S. aureus NCTC8325. The genetic diversity appears to be important in determining evolutionary relationships, following the epidemiology of specific bacterial isolates. Furthermore, an overview about invasiveness and virulence of some isolates especially in hospitals can be assessed (Onasanya et al., 2003). Little studies were estimated the S. aureus resistant to methicillin according to published researches in Iraq. A variable genetic relatedness of some clinically isolated S. aureus isolates was documented (Othman et al., 2014).

References:

- Al-Dahbi AM and Al-Mathkhury HJ. Distribution of Methicillin Resistant *Staphylococcus aureus* in

Iraqi patients and Healthcare Workers. Iraq J Sci 2013; 54(2): 293-300.

- Al-Mayahie SMG, Al-Hamashee HTR, Hameed MH. Prevalence and Antimicrobial Susceptibility of Methicillin-Resistant *Staphylococcus aureus* (MRSA) from Outpatients with Chronic Rhinosinusitis in Al-Kut/Wasit Province/Iraq. Bacteriol Parasitol 2015; 6(3): 1-6.
- AL-Ruaily MA and Khalil OM. Detection of (*mecA*)gene in methicillin resistant *Staphylococcus aureus* (MRSA) at Prince A/Rhman Sidery Hospital, Al-Jouf, Saudi Arabia.J Med Gen 2011; 3(3): 41-5.
- Bauer AW, Kirby WM, Sherris JC, Tyrck M. Antibiotic susceptibility testing by standardized single method. Am J Clin Pathol 1966; 45(4): 493-6.
- Clinical Laboratory Standards Institutes. Performance standards for antimicrobials susceptibility testing; Nineteenth informational supplement, 2009. M100-S19, 29(3).
- Degaim ZD, Shani WS, Hamim SS. Virulence factors of Methicillin Resistant *Staphylococcus aureus* (MRSA) isolated from burn patients. Int J Curr Micobiol App Sci 2015; 4(7):898-906.
 Ghasemiam A and Mirzaee M. Methicillin Resistant *Staphylococcus aureus* (MRSA) Strains and the Staphylococcal Cassette Chromosome *mec* Types in Iran. Infect Epidemiol Med 2016; 2(3): 31-4.
- Hall TA. Bioedit: a user-friendly biological sequence alignment editor and analysis program for windows 95/98nt. Nucl Acids Symb ser 1999; 41: 95-8.
- Harley JP and Prescott LM. Laboratory Exercises in Microbiology. 5th ed. New York. McGraw-Hill; 2002.
- Hussain NR. Prevalent Genotypes of *Staphylococcus aureus* Strains Isolated From Healthcare Workers in Duhok City, Kurdistan Region, Iraq. Int J Infect 2016; 3(2): e35375.
- Jarvis WR, Schlosser J, Chinn RY, Tweeten S, Jackson M. National prevalence of methicillinresistant *Staphylococcus aureus* in inpatients at US health care facilities. Am J Infect Control 2007; 35:631-37.
- Jensen SO, Lyon BR. Genetic of Antimicrobial Resistance in *Staphylococcus aureus*. Fut Microbiol 2009; 4(5):565-82.

Website: http://jsci.utq.edu.iq

Volume 6, Number 1, December 2016

- Jonas D, Speck M, Daschner FD, Grundmann H. Rabid PCR-based identification of methicillinresistant *Staphylococcus aureus* from screening swabs. J Clin Microbiol 2002; 40(5):1821-23.
- Kumar S, Stecher G, Tamura K. MEGA7: Molecular Evolutionary Genetics Analysis version 7.0 for bigger datasets. Mol Biol and Evol 2016; 33:1870-74.
- Memmi, G Filipe SR, Pinho, MG., Fu Z. Cheung, A. *Staphylococcus aureus* PBP4 is essential for betalactam resistance in community-acquired methicillin-resistant strains. Antimicrob Agents Chemother 2008; 52: 3955-66.
- Onasanya A, Mignouna H, Thottappilly G. Genetic fingerprinting and phylogenetic diversity of *Staphylococcus aureus* isolates from Nigeria. Afric J Bio 2003; 2 (8):246-50.

- Othman HE, Merza NS Jubrael JMS. Nucleotide Sequence Analysis of Methicillin Resistance *Staphylococcus aureus* in Kurdistan Region-Iraq. J Univ. Zak 2014; 2A(1): 1-12.

- Rahimi F and Karimi S. Characterization of strains isolated from poultry in Iran. Arch Clin Infect Dis 2015; 10(4):1-9.
- -Rahimi F, Katouli M, Pourshafie MR. Characterization of hospital and community-acquired methicillinresistant *Staphylococcus aureus* in Tahran, Iran. Iran J Med Microbiol 2014;63(6):796-804.
- Schito GC. The importance of the development of antibiotic resistance in *Staphylococcus aureus*. Clin Microbiol Infect 2006; 12(S1):3-8.
- Shanson DC. Antibiotic-resistant *Staphylococcus aureus*. J Hosp. Infet 1981; 2(1): 11-36.
- Wannet W, Heck M, Pluister G, Spalburg E, De Neeling, AJ. Panton-valentine leucocidin positive MRSA in 2003: the Duch situation. Eur. Surv 2004; 9:28-9.

Email: utjsci@utq.edu.iq